Experimental study of efficacy of intralamellar keratoplasty with corneal stromal substitute developed from decellularized porcine cornea

N.V. Pasyechnikova, Dr. Sc. (Med), Prof.

B.M. Cogan, Cand Sc (Med)

S.G. Kolomiichuk, Research Fellow

Filatov Institute of Eye Diseases and Tissue Therapy of the NAMS of Ukraine, Odessa, Ukraine

E-mail: koganb@ukr.net

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Backrgound: Enhancing methods for decellularization of animal corneas for the development of bioengineered corneal models with the following keratoplasty makes it possible to overcome the shortage of human donor material.

Purpose: To investigate the clinical efficacy of intralamellar keratoplasty (IKP) with corneal stromal substitutes (CSSs) developed using different decellularization methods in the animal study.

Materials and Methods: Corneas were excised from enucleated pig eyes, and, to develop acellular CSSs, five methods of decelluarization of porcine corneas were used that were different in the time and conditions of incubation with detergents (0.5% sodium dodecyl sulphate or TRITON X-100) and proteolytic enzymes (like 0.1% papain). In addition, an ultrasonic apparatus of type CD 3800 A was used to further improve the removal of cells and acellular corneal debris. A total of 40 Chinchilla rabbits were used in the animal study. They were divided into 5 groups with different CSS versions. In each rabbit, the monocular IKP was carried out with one of the five CSS versions. Postoperatively, antimicrobial and antiinflammatory therapy was instituted. Animals were examined every other day for 30 days postoperative. This included corneal fluorescein staining. In addition, the following was assessed ophthalmoscopically: conjunctival discharge, conjunctival hyperemia, state of the CSS, state of the host cornea, and, at late follow-up examinations, graft acceptance and whether graft-versus-host disease was present. Results: In rabbit eyes that underwent IKP with a version 4 CSS, the clinical scores for conjunctival discharge, corneal graft edema, inflammatory infiltration, corneal graft opacity and corneal fluorescein staining (0.13 points) were statistically significantly lower than in eyes that underwent IKP with other CSS versions. Location of inflammation in the cornea was found to be paracentral in eyes that improved clinically after IKP with version 1, 2, 3 or 5 CSS, and tended to be central (0.38 points) in those that underwent IKP with a version 4 CSS.

Conclusion: The least apparent reaction was noted in the rabbit eyes that underwent IKP with a version 4 CSS, which makes this CSS promising for further investigations in pre-clinical and clinical studies.

Introduction

Today, in spite of advances in our knowledge of the etiopathogenesis of different anterior eye disorders, corneal disease is a major cause of loss of vision. For many cases with a severe clinical form of corneal injury, surgical treatment, namely, keratoplasty, is therefore the only effective option [1-3].

Because the application of such a radical treatment (as keratoplasty) is hampered by the risk of graft rejection in antigen incompatibility between graft and host, and by low availability of adequate donor material, developing alternatives to human donor cornea is of great importance [1, 4-13].

In the last decade, xenotransplantation (i.e., the transplantation of animal organs, tissue and cells into human beings) has been used to overcome the shortage of human donor material. The use of porcine organs (especially, islets of Langerhans) for transplantation appears to be the most promising option, given in particular, that the pig most closely resembles humans with regard to antigenic composition of tissues [6, 8, 14].

In this connection, it seems important to consider the potential for using porcine cornea as a xenograft in humans.

We should give attention to the fact that after decellularization, the xenograft should be similar to the native human cornea with regard to immunological characteristics, biocompatibility, mechanical strength and transparency, thus creating an optimal micro-

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environment for subsequent migration of stromal and epithelial cells [8, 10, 15-17].

Therefore, developing and enhancing methods for decellularization of donor corneas for the development of bioengineered corneal models with the following keratoplasty is a task to be tackled by experimental and clinical ophthalmologists [5-7, 11, 16-17].

The purpose of this experimental animal study was to investigate the clinical efficacy of intralamellar keratoplasty with corneal stromal substitutes (CSS) developed using different decellularization methods.

Materials and Methods

Corneas were excised from enucleated pig eyes, and, to develop acellular CSSs, five methods of decelluarization of pig corneas were used that were different in the time and conditions of incubation with detergents (0.5% sodium dodecyl sulphate or TRITON X-100) and proteolytic enzymes (like 0.1% papain) [6]. In addition, an ultrasonic apparatus of type CD 3800 A (power, 50 W) was used to further improve the removal of cells and acellular corneal debris.

A total of 40 Chinchilla rabbits (80 eyes; weight, 2.5–3 kg) were used in the experimental study. They were divided into 5 groups with different CSS versions.

Preoperatively, animals were anesthetized with thiopental sodium (1 g/kg, intravenously). In each rabbit, the monocular IKP was carried out [1, 18] with one of the five CSS versions in the operating room setting. Postoperatively, antimicrobial and anti-inflammatory therapy was instituted. Animals were examined every other day for 30 days postoperative. This included fluorescein staining to assess corneal epithelial status. In addition, the following was assessed ophthalmoscopically: conjunctival discharge, conjunctival hyperemia, state of the CSS, state of the host cornea, and, at late followup examinations, graft acceptance and whether graftversus-host disease was present [6]. The following items related signs of to were scored 0 to 3: conjunctival discharge (0, no discharge; 1, mild mucous discharge; 2, severe mucous discharge; 3, purulent mucous discharge), conjunctival hyperemia (0, pale pink conjunctiva as per physiological norm; 1, mild bulbar conjunctival hyperemia; 2, moderate bulbar conjunctival hyperemia; 3, severe bulbar conjunctival hyperemia), corneal graft edema (0, no edema and the whole cornea is transparent; 1, local corneal epithelial edema at inflammation site; 2, local epithelial edema with involvement of superficial stromal layers; 3, local edema involving both superficial and deep stromal layers), corneal fluorescein staining (0, no local staining; 1, punctate corneal staining; 2, less than 3 mm² area; 3, more than 3 mm2 area). Inflammatory infiltration was scored 0 to 4 (0, no infiltration; 1, one to three punctate subepithelial infiltrates; 2, multiple punctate subepithelial infiltrates (n > 3); 3, multiple subepithelial infiltrates of 1 mm at least; 4, local infiltration involving both superficial and deep stromal layers). Corneal graft opacity was scored 0 to 1 (0 = no opacity; 1 = presenceof opacity). Location of inflammation in the cornea was scored 0 to 1 (1, central; 2, paracentral).

Animals were operated on and euthanized (at 1 month postoperatively) in accordance with the General Ethical

Principles for Animal Experiments approved by the Third National Congress on Bioethics (Kyiv, 2007), and the European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes (European Treaty series no. 123).

Box plots were used to display the clinical rabbit eye scores after IKP with different group-related CSS versions (Nos.1-5; Figs. 1-7).

Statistical analyses were conducted using Statistica 10.0 (StatSoft, Tulsa, OK, USA) software. The Kruskal Wallis test was used to compare the clinical scores at 1 month after IKP with a CSS within the groups. Pairwise comparisons among the groups were conducted using the Mann-Whitney test.

Results and Discussion

A Kruskal-Wallis test showed that there were statistically significant differences in the scores of conjunctival discharge, conjunctival hyperemia, corneal graft edema, corneal graft opacity, and location of corneal inflammation among the groups after IKP with a group-related CSS (Table 1).

A Mann-Whitney test showed that, in group 1, the mean conjunctival discharge, conjunctival hyperemia and corneal graft edema scores were 4.3, 4.3 and 3.6-fold higher, respectively, than in group 2 (Figs. 1-7).

There were statistically significant differences in the scores of conjunctival discharge (p = 0.01), conjunctival hyperemia (p = 0.01), and corneal graft edema (p = 0.005) between group 1 and group 2.

In group 3, the mean conjunctival discharge, conjunctival hyperemia and corneal graft edema scores were 6.5, 6.5 and 3.6-fold lower, respectively, than in group 1 (Figs. 1-7). There were statistically significant differences in the mean scores of conjunctival discharge (p = 0.01), conjunctival hyperemia (p = 0.01), and corneal graft edema (p = 0.005) between group 3 and group 1.

It should be noted that the mean scores of conjunctival discharge, conjunctival hyperemia, and corneal graft edema in group 4 were very low (0.13 points) (Figs. 1-7). In this group, the mean conjunctival discharge, conjunctival hyperemia and corneal graft edema scores were 12.5, 12.5 and 10.6-fold lower, respectively, than in group 1 (p = 0.001) (Figs. 1-7).

In group 5, the mean conjunctival discharge, conjunctival hyperemia and corneal graft edema scores were 6.5 (p = 0.001), 6.5 (p = 0.001) and 3.6-fold (p = 0.005) lower, respectively, than in group 1 (Figs. 1-7).

Although there were no statistically significant differences in the mean scores of conjunctival discharge, conjunctival hyperemia, and corneal graft edema between group 2 and group 3, the mean scores of conjunctival discharge and conjunctival hyperemia in the latter group were 1.5 and 1.5-fold, lower, respectively, than in the former group (Figs. 1-7).

Eyes that received IKP with version 4 CSS showed 2.9-fold lower scores for conjunctival discharge, conjunctival hyperemia and corneal graft edema than those that received IKP with version 2 CSS, but these differences were also not significant (Figs. 1-7).

In group 5, the mean conjunctival discharge and conjunctival hyperemia scores were 1.5-fold lower than in group 2, while the mean corneal graft edema score was the same in both groups (Figs. 1-7).

In group 5, the mean inflammatory infiltration score was 2.9-fold lower, whereas the mean fluorescein staining and corneal graft opacity scores were 1.5 and 1.9-fold higher, respectively, than in group 3. No difference in conjunctival discharge, conjunctival hyperemia, and corneal graft edema was observed between the two groups (p > 0.05) (Figs. 1-7).

In group 4, the mean conjunctival discharge, conjunctival hyperemia and corneal graft edema scores were 1.9, 1.9 and 2.9-fold lower, respectively, than in group 5 (Figs. 1-7).

The mean corneal graft opacity score was better in eyes that received IKP with version 3 or 4 CSS (0.13 points) than in those that received IKP with version 2 or 5 CSS (0.25 points) or version 1 CSS (0.75 points). That is, in group 2 and group 5, the mean corneal graft opacity scores were 3-fold lower, than in group 1, whereas in group 3 and group 4, they were 5.8-fold lower, than in group 1.

The mean corneal graft opacity scores in eyes that received IKP with version 3 or 4 CSS were 1.9-fold lower than in those that IKP with version 2 or 5 CSS.

The differences in mean corneal graft opacity scores between group 1 and group 3, and between group 1 and group 4, were statistically significant (p < 0.015 and p < 0.015, respectively).

Both Kruskal-Wallis test (Table 1) and Mann-Whitney test revealed no significant differences in the scores for inflammatory infiltration and corneal fluorescein staining among the five groups representing different CSS versions.

Eyes that received IKP with version 1, 4 or 5 CSS showed the best mean scores for inflammatory infiltration (0.13 points), and eyes that received IKP with version 4 CSS showed the best mean scores for corneal fluorescein staining (0.13 points) (Figs. 1-7).

The inflammation location was paracentral in group 1 (2.63 points), group 2 (2.75 points), group 3 (2.87 points) and group 5 (2.75 points), whereas in group 4, the inflammation location score was 0.38 due to low extent of pathologic changes (Figs. 1-7).

There was a significant intergroup difference in inflammation location score between the following pairs of groups: group 1 and group 4 (p = 0.003), group 2 and group 4 (p = 0.002), group 3 and group 4 (p = 0.001) and group 4 and group 5 (p = 0.002).

Although numerous reports have been published on decellularization methods, many of them are related to experimental in vitro studies [9,10,19], and the CSSs developed in these studies do not always meet the requirements for mechanical strength, transparency and biocompatibility. Today, decellularized corneas are considered a better choice for keratoplasty than synthetic or semi-synthetic substitutes [5,9,10] because of higher mechanical indices and their potential for preserving the

native matrix ultrastructure and intrinsic biological cues like growth factors [20]. The supply of human donor material is, however, limited by legal and ethical issues. If a currently available alternative to human donor cornea (like any xenograft development) [10,17,21] is to qualify for clinical use, a requirement which must be met is standardization of the most suitable decellularization methods. Moreover, potential cytotoxicity of various compounds used for decellularization of the cornea makes the requirements for decellularization conditions stricter.

Although promising results have been reported by Pasyechnikova et al [18], Shafiq [22], Nasinnik [4] and Wilson [20] for the use of decellularized human corneal stromal pieces, this approach is limited by the issues related to donor material harvesting.

The use of physical methods like high-hydrostatic pressurization for corneal tissue engineering has been demonstrated to provide decellularized cornea with good optical quality and without ultrastractural changes [23]; however, it may disrupt ECM ultrastructure [24]. In addition, post-pressurization immersion of decellularized cornea in glycerol for 1 hour resulted in restoration of the corneal optical properties to those of native corneas [25].

As opposed to Lee et al [26] and Yoeruek et al [27], we used 1) a combination of proteolytic enzymes with ultrasound pretreatment of corneal stroma for improved decellularization of porcine cornea, and 2) intralamellar keratoplasty to assess the developed corneal stromal substitutes.

In our studies, the use of xenotransplantation and particular decellularization methods enabled us to solve the issues related to donor material harvesting and biocompatibility.

We performed IKP with five different versions of CSS in eyes of experimental animals, and thereafter compared clinical rabbit eye indices among the groups with different CSS versions. This enabled us to determine a decelluarization method for the development of CSS that is adequately strong, relatively simple to process, and has a low production cost compared with prototypes.

Conclusion

The lowest clinical scores for conjunctival discharge, corneal graft edema, inflammatory infiltration, corneal graft opacity and corneal fluorescein staining (0.13 points) were noted in rabbit eyes that underwent intralamellar keratoplasty with a version 4 corneal stromal substitute. Location of inflammation in the cornea was found to be paracentral in eyes that improved clinically after IKP with version 1, 2, 3 or 5 CSS, and tended to be central (0.38 points) in those that underwent IKP with version 4 CSS. Therefore, the least apparent reaction was noted in the rabbit eyes that underwent IKP with a version 4 CSS, which makes this CSS promising for further investigations in pre-clinical and clinical studies.

Table 1. Comparison of clinical indices of rabbit eyes after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute (n=8 animals/eyes for each version)

Clinical indices	Stat index	Corneal stromal substitute version					р
		1	2	3	4	5	
Conjunctival discharge	Average rank	35.00	19.06	16.88	14.69	16.88	0.001
Conjunctival hyperemia	Average rank	35.00	19.06	16.88	14.69	16.88	0.001
Corneal graft edema	Average rank	33.38	18.44	18.44	13.81	18.44	0.002
Inflammatory infiltration	Average rank	18.50	23.50	23.50	18.50	18.50	0.500
Corneal fluorescein staining	Average rank	23.25	20.25	20.25	17.88	20.88	0.825
Corneal graft opacity	Average rank	29.50	19.50	17.00	17.00	19.50	0.041
Corneal infiltrate location	Average rank	21.81	23.88	25.94	7.00	23.88	0.001

Note. P values were calculated using a Kruskal-Wallis test for multiple comparisons

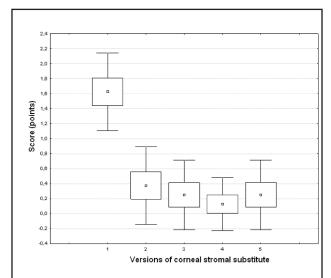


Figure 1. Conjunctival discharge scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

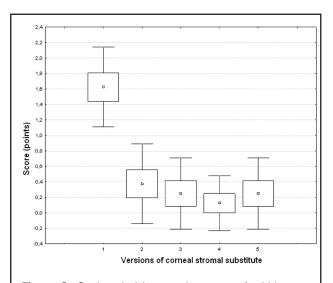


Figure 2. Conjunctival hyperemia scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

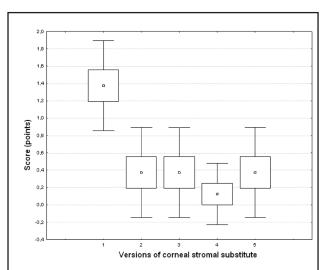


Figure 3. Corneal graft edema scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

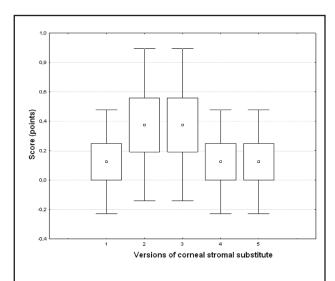


Figure 4. Inflammatory infiltration scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

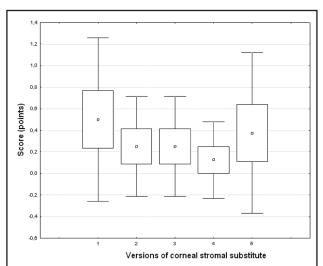


Figure 5. Corneal fluorescein staining scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

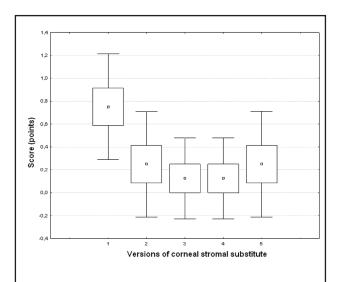


Figure 6. Corneal graft opacity scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\rm o}$, mean; , standard error; $^{\perp}$, standard deviation

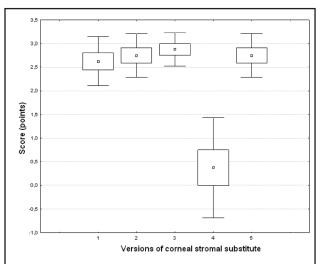


Figure 7. Corneal infiltrate location scores of rabbit eyes at day 30 after intralamellar keratoplasty with different versions (1 to 5) of corneal stromal substitute. $^{\circ}$, mean; , standard error; $^{\perp}$, standard deviation

References

- Dushin NV, Frolov MA, Gonchar PA. [Keratoplasty for optical, refractive, curative and esthetic indications in the treatment of ocular disorders: a tutorial]. Moscow: RUDN; 2008. Russian
- Kaminski SL, Biowski R, Lukas JR, et al. Corneal sensitivity 10 years after epikeratoplasty. J Refract Surg. 2002 Nov-Dec;18(6):731-6
- Whitcher JP, Srinivasan M, Upadhyay MP Corneal blindness: a global perspective. Bull WHO. 2001; 79: 214– 21
- Nasinnik IO. [Experimental substantiation for the use of corneal allograft]. [Abstract of Cand Sc (Med) Thesis]. Odessa: Filatov Institute of Eye Disease and Tissue Therapy; 2013. 19 p. Russian
- Pasyechnikova NV, Vit VV, Leus NF, et al. [Clinical and histological results of experimental use of implants based on cross-linked collagen as donor cornea analogue]. Oftalmol Zh. 2011;4:58-60. Russian.
- 6. Pasyechnikova NV, Kogan BM, Gladush TI, et al. [The study of efficacy of in layer transplantation of heterogenous transplant of the cornea equivalent in the experiment]. Oftalmol Zh. 2014;2:64-70. Russian.
- 7. Choi JS, Williams JK, Greven M et al. Bioengineering endothelialized neo—corneas using donor—derived corneal endothelial cells and decellularized corneal stroma. Biomaterials. 2010 Sep;31(26):6738-45
- 8. Cox A, Zhong R. Current advances in xenotransplantation. Hepatobiliary Pancreat Dis Int. 2005 Nov;4(4):490-4
- A simple, cross—linked collagen tissue substitute for corneal implantation. Liu Y, Gan L, Carlsson DJ, et al. Invest Ophthalmol Vis Sci. 2006 May;47(5):1869-75
- Ponce Mórquez S, Marthnez VS, McIntosh Ambrose W, et al. Decellularization of bovine corneas for tissue engineering applications. Acta Biomater. 2009 Jul;5(6):1839-47
- Pasyechnikova N., Vit V, Leus N, et al. Collagen-Based Bioengineered Substitutes of Donor Corneal Allograft Implantation: Assessment and Hypotheses. Med Hypothesis Discov Innov Ophthalmol. 2012 Spring; 1(1): 10–13
- 12. Rafat M, Li F, Fagerholm P, et al. PEG-stabilized carbodiimide crosslinked collagen—chitosanhydrogels for corneal tissue engineering. Biomaterials. 2008;29(29):3960—72
- 13. Thompson RW Jr, Price MO, Bowers PJ, Price FW Jr. Long—term graft survival after penetrating keratoplasty. Ophthalmology. 2003 Jul;110(7):1396-402
- Bartholomew LR, Pang DX, Sam DA, Cavender C. Ultrasound biomicroscopy of globes from young adult pigs. Am J Vet Res. 1997 Sep;58(9):942-8

- 15. Duan X, Sheardown H. Dendrimer crosslinked collagen as a corneal tissue engineering scaffold: mechanical properties and corneal epithelial cell interaction. Biomaterials. 2006 Sep;27(26):4608-17
- Gilbert TW, Sellaro TL, Badylak SF. Decellularization of tissues and organs. Biomaterials. 2006 Jul;27(19):3675-83
- 17. Wu Z, Zhou Y, Li N, et al. The use of phospholipase A2 to prepare acellular porcine corneal stroma as a tissue engineering scaffold. Biomaterials. 2009;30(21):3513–22
- 18. Pasyechnikova NV, Vit VV, Leus NF, et al. [The reaction of the rabbit cornea after intralamellar transplantation of the acellular stroma of the human cornea]. Oftalmol Zh. 2011;1:57-60. Russian.
- 19. Yoeruek E, Bayyoud T, Maurus C, et al. Decellularization of porcine corneas and repopulation with human corneal cells for tissue—engineered xenografts. Acta Ophthalmol. 2012;90: e125-31
- Wilson SL, Sidney LE, Dunphy SE, et al. Corneal Decellularization: A Method of Recycling Unsuitable Donor Tissue for Clinical Translation? Curr Eye Res. 2016 Jun;41(6):769-82
- Lu Y, Yao QK, Feng B, et al. Characterization of a Hydrogel Derived from Decellularized Corneal Extracellular Matrix.
 J Biomaterials and Tissue Engineering. 5: 951-60
- 22. Shafiq MA, Gemeinhart RA, Yue BY, Djalilian AR. Decellularized human cornea for reconstructing the corneal epithelium and anterior stroma. Tissue Eng Part C Methods. 2012 May;18(5):340-8
- 23. Hashimoto Y, Sasaki S, Hattori S, et al. Ultrastructural analysis of the decellularized cornea after interlamellar keratoplasty and microkeratome-assisted anterior lamellar keratoplasty in a rabbit model. Sci Rep. 2016; 6: 27734
- 24. Crapo PM, Gilbert TW, Badylak SF. An overview of tissue and whole organ decellularization processes. Biomaterials. 2011 Apr;32(12):3233-43
- 25. Hashimoto Y, Funamoto S, Sasaki S, et al. Preparation and characterization of decellularized cornea using high-hydrostatic pressurization for corneal tissue engineering. Biomaterials. 2010;31(14):3941-8
- 26. Lee W, Miyagawa Y, Long C, et al. A comparison of three methods of decellularization of pig corneas to reduce immunogenicity. Int J Ophthalmol. 2014; 7(4): 587–93.
- 27. Yoeruek E, Bayyoud T, Maurus C, et al. Reconstruction of corneal stroma with decellularized porcine xenografts in a rabbit model. Acta Ophthalmol. 2012;90: e206-10